

Assessment of Foodborne Disease Hazards in Beverages Consumed in Nigeria: A Systematic Literature Review

David O. Oduori,^{1,2,i} Emmah Kwoba,¹ Lian Thomas,^{1,3} Delia Grace,^{1,4} and Florence Mutua^{1,ii}

Abstract

Risk assessment is a formal process of identifying hazards and assessing the risk associated with them (risk is a combination of the severity of illness and the probability of occurrence). This review highlights foodborne disease hazards reported in beverages consumed in Nigeria for the period between 2000 and 2020. Based on a preregistered protocol and search syntax, studies were retrieved from the PubMed, Google Scholar, and ScienceDirect databases. Rayyan QCRI software was used to screen the articles. Data were then extracted from the included full-text articles, into a standardized excel workbook. A total of 18,762 articles were identified, from which 126 were included in the final analyses. The common beverages studied were sachet water (14.9%), borehole/well water (13.9%), cereal-based beverages (12.1%), raw/fresh milk (8.3%) and *nono/nunu*, which is a fermented milk-cereal beverage (7.2%). Sufficient data were available to undertake pooled prevalence estimates for some hazards within select beverages and revealed contamination rates for *Staphylococcus* spp. in raw/fresh milk, 12.3% (95% CI 6.3–20.0); *Salmonella* spp. in borehole/well water, 19.8% (95% CI 13.1–27.4); *Klebsiella* spp. in sachet water, 40.0% (95% CI 12.4–71.7); *Staphylococcus* spp. in *nono/nunu*, 32.6% (95% CI 14.7–53.8), and *Escherichia* spp. in *nono/nunu*, 30.7% (95% CI 21.9–40.2). Heterogeneity was present in the aggregate summary estimates. This review has highlighted the presence of several hazards of high importance to public health in commonly consumed beverages in Nigeria. The data presented here provide an entry point for future quantitative risk assessments both to determine the level of exposure of the community to these hazards and also for the identification of the most effective mitigation strategies to reduce these risks and improve health outcomes in Nigeria.

Keywords: foodborne disease, hazards, beverages, Nigeria

Introduction

GLOBALLY, 420,000 DEATHS AND 600 million illnesses were attributed to foodborne diseases caused by infectious agents in 2010 (Havelaar *et al.*, 2015). The highest populous affected was from Africa followed by Southeast Asia, with diarrheal disease agents being implicated as the major contributor, responsible for over 50% of the deaths (Muller and Krawinkel, 2005; Havelaar *et al.*, 2015). Consequently, the global foodborne disease burden is estimated at 33 million Disability Adjusted Life Years (DALYs), 40%

of this is borne by children younger than 5 years (Muller and Krawinkel, 2005; Havelaar *et al.*, 2015). Contamination of food by heavy metals is responsible for an estimated 1 million illnesses, over 56,000 mortalities, and a DALY metric of 9 million globally. This estimate considers chemical contamination of food by arsenic, methylmercury, lead, and cadmium only (Gibb *et al.*, 2019).

Food safety has been flagged as a significant barrier toward social and economic development and in the attainment of Sustainable Development Goals 1–3, “No Poverty,” “Zero Hunger,” and “Good Health and Well-being” (Havelaar

¹International Livestock Research Institute, Nairobi, Kenya.

²Department of Animal Health and Production, Maasai Mara University, Narok, Kenya.

³Institute of Infection Veterinary & Ecological Sciences, University of Liverpool, Wirral, United Kingdom.

⁴Natural Resource Institute, University of Greenwich, Kent, United Kingdom.

ⁱORCID ID (<https://orcid.org/0000-0002-7664-6938>).

ⁱⁱORCID ID (<https://orcid.org/0000-0002-1007-5511>).

et al., 2015). The inverse relationship between a decline in economic prosperity and foodborne illness in developing societies suggests that addressing food safety may positively contribute to economic gains (Akhtar *et al.*, 2014). Foodborne disease costs developing countries at least \$100 million a year (Jaffee *et al.*, 2020).

Subregion “AFR D,” to which Nigeria belongs, was found to have the highest burden of foodborne illness: 1276 (459–2263) DALYs per 100,000 population (Havelaar *et al.*, 2015). The frequency of illnesses related to the consumption of contaminated food could imply the lack of efficient food safety control systems (Mensah *et al.*, 2012; Paudyal *et al.*, 2017). For Nigeria, this assertion is supported by outputs from anthropometric surveys that report a stunting and malnutrition prevalence of 36.8% and 8.9%, respectively, in children younger than 5 years (NPC and ICF, 2019). Bacterial and parasitic diseases are known risk factors for malnutrition in developing countries (Muller and Krawinkel, 2005). The burden due to aflatoxins has also been shown to be high in some African countries, including Nigeria (Havelaar *et al.*, 2015).

Interventions to address hazards in foods are urgently needed. Determining what strategies can be used, alongside other key health burdens across society, should, however, be done in a rational and, where possible, objective manner. It is not only the burden of disease attributable to pathogens or products that should be considered, but also the cost-effectiveness or net economic benefit of potential control interventions. The initial process in such systematic prioritization processes is a risk analysis where, in a stepwise manner, hazards and the population at risk are identified (risk ranking), a quantification (or semiquantitative evaluation) of the risk posed by the hazards is made (risk assessment), risk mitigation steps are identified (risk management), and finally, risk communication is undertaken. This study contributes to the first steps of risk ranking and risk assessment by identifying foodborne hazards of potential risk to Nigerian consumers. Many foodborne pathogens can be found in common beverages, with water being a well-known example of a beverage with a high burden of disease attributable to the pathogens it may carry. Beverages consumed in Nigeria have been reported to harbor various hazards and are thus a public health concern.

A Systematic Literature Review (SLR) was undertaken to establish the current evidence on foodborne hazard occurrence in a myriad of beverages consumed in Nigeria between the year 2000 and the year 2020. The information will inform further risk assessment work in Nigeria. It will assist in determining the riskiest beverage products, potential mitigation activities, and their impacts. This review, therefore, highlights hazards associated with commonly consumed beverages in Nigeria. The findings will inform the scope of further risk assessment work in foods in formal and informal markets in Nigeria, and other countries with similar situations.

Materials and Methods

Protocol development

A protocol to guide the SLR process was developed and registered in PROSPERO and is searchable in <https://www.crd.york.ac.uk/prosperto/> using CRD42020184768 as the registration number.

Literature search

Searches were done in three search engines, namely PubMed, Science Direct, and Google Scholar. The search was set to identify articles published in the period 2000–2020. An initial syntax covering most beverages and safety terms was developed for PubMed using Medical Subject Headings (MeSH) terms and Boolean search operators (Supplementary Data S1). Since ScienceDirect has a syntax string limit of 8 Boolean expressions, therefore a series of 40 short syntaxes, including all the beverage and safety terms in the PubMed syntax, was developed. These articles were exported to the Mendeley reference manager from where duplicates were identified and removed. The resulting unique file was treated as a single output from ScienceDirect. Similarly, for Google Scholar, which has a character limit of 256, a series of 20 syntaxes was developed to include all the beverage and safety terms in the syntax used in PubMed. For each search output, the first 300 hits were considered for the review (Haddaway *et al.*, 2015) and were exported to Mendeley via its Web Importer function. All the search outputs were pooled in the Mendeley reference manager, and the duplicates were identified and removed, resulting in a unique Google Scholar file.

Article screening

This review included observational studies published between 2000 and 2020. Only studies conducted in Nigeria and those published in English were included in the review. Studies with experimental design, on water quality/safety not associated with drinking water, antimicrobial resistance, and those not considering biological or chemical hazards associated with beverage consumption were excluded.

The screening process was facilitated by the Rayyan QCRI (<https://rayyan.qcri.org/>), a web-based and mobile-based application that is used for screening articles in SLRs (Ouzani *et al.*, 2016). The search outputs from the three databases were exported (directly in the case of PubMed and from Mendeley in the case of ScienceDirect and Google Scholar) to the software where any duplicates were identified and subsequently resolved. Publication titles and abstracts were then screened against the inclusion and exclusion criteria as specified in the preregistered protocol. Four reviewers participated in the process. The screening was done independently by two reviewers, Reviewers 1 and 2. Articles were then subjected to full-text screening against the predetermined criteria and reasons for the exclusion provided. Articles that were found acceptable after the full-text screening were considered for data extraction. Any discordance in the classification of articles by Reviewers 1 and 2 was resolved by Reviewers 3 and 4, and this applied for both abstract and full article screening. For quality control, 5% of the included and 5% of the excluded articles were reviewed by Reviewers 3 and 4. Additional articles were identified through screening of the reference section of the included publications. The output of the screening process was then reported according to the PRISMA 2009 flow diagram guidelines (Moher *et al.*, 2015).

Quality assessment and data extraction

Data extraction and quality assessment happened concurrently. Quality assessment of individual articles was based on the Cochrane assessment of bias (Higgins *et al.*, 2021).

The articles were classified as being of good, medium, and poor quality. Articles that had an unbiased selection of subjects, where the methods were judged to be scientifically sound, and appropriate data analysis had been conducted with complete and accurate results were judged as good quality. Medium-quality articles had acknowledged and accounted for selection bias of subjects, limitations in data analysis, understandable methods, and valid results. Articles that did not acknowledge selection bias of subjects had incomplete and inaccurate methods, inappropriate data analysis, and incomplete results judged to be of poor quality were excluded from the study, and data were not extracted from them.

A data extraction template in Microsoft Excel was developed and piloted. The template was designed to capture the following data: article identifiers (article ID, the title of the publication, authors, year of publication), type of study, geographical locality, name of the beverage, name and category of hazard, point of sampling, sampling technique,

specific laboratory test used, number of samples analyzed, number positive for the hazard, and raw data on the concentration of hazards. For review articles, data were not extracted directly from them but from the primary data, articles cited.

Data management and analysis

The data were cleaned and validated to check for any errors and omissions. They were analyzed using the R platform for statistical computing software (R Core Team, 2018). Data on type of beverage, type of study, category, and specific hazards, point and method of sampling, and laboratory test used were summarized as proportions and presented as tables and figures. A random-effects model was selected for estimating pooled prevalence due to the interstudy differences (Bown and Sutton, 2010). To ascertain stronger statistical inference, pooled prevalence was estimated for hazard/beverage combinations where $5 \geq$ good-quality studies were available

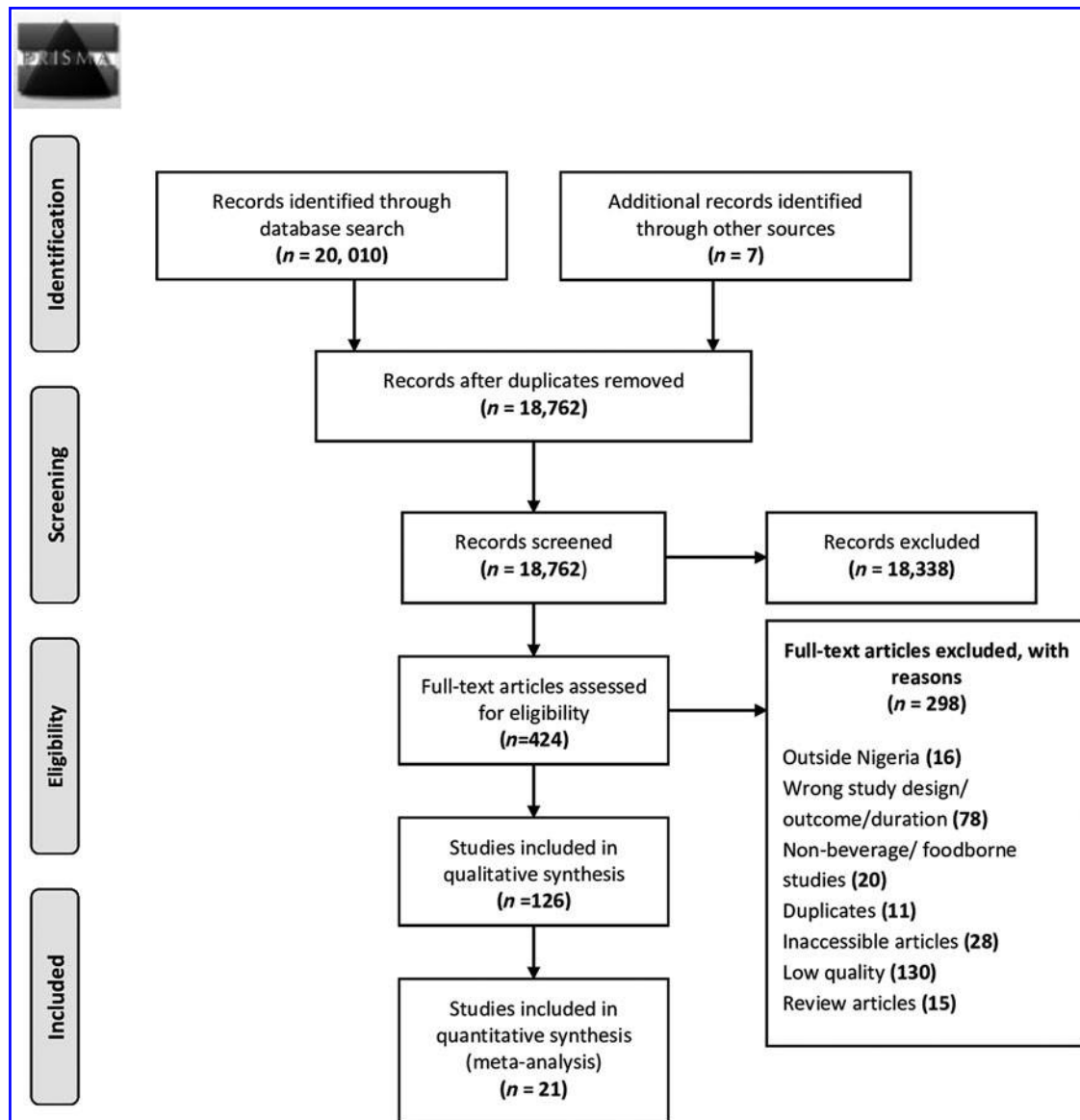


FIG. 1. PRISMA 2009 flow diagram occurrence of FBD hazards in beverages consumed in Nigeria. FBD, foodborne disease. From Moher *et al.* (2009). For more information, visit www.prisma-statement.org

(Jackson and Turner, 2017). Forest plots and summary tables including estimates of heterogeneity were generated using the metaviz package of R software. Heterogeneity thresholds were derived from Deeks *et al.* (2019), where I^2 of 0–40% represented low heterogeneity or homogeneity.

Results

Database search output and screening

A total of 20,100 articles were retrieved from the 3 databases: PubMed (1315), ScienceDirect (14,406), and Google Scholar (4289). Seven additional articles were identified from review articles. A total of 1255 duplicates were removed (either in Mendeley or in Rayyan QCRI). After title and abstract screening, 18,338 articles were excluded, resulting in 424 articles that were subjected to full-text screening. Two hundred ninety-eight articles were excluded at the stage of full article review on the following basis: low quality (130), wrong study design or outcome (77), inaccessible articles where the full text was not available (28), nonbeverage or nonfoodborne studies (20), articles with a geographical focus outside Nigeria (16), review articles (15), additional duplicates not previously identified (11), and articles outside the time frame of interest (1). A total of 126 articles were thus included in the qualitative analyses, of which 21 articles were included in pooled prevalence analysis. The PRISMA 2009 flowchart is given in Figure 1.

Characteristics of articles included in the qualitative synthesis

The 126 articles included in the qualitative analyses resulted in a total of 787 records. Among the 126 articles, 73.8% had some details lacking in the methods section, but bias in the selection of subjects and limitations in data analysis had been acknowledged and the articles were therefore deemed to be of medium quality. Only 26.2% of the articles were judged to be of good quality, as they had an unbiased selection of subjects, scientifically sound study methods, appropriate data analysis, and complete and accurate results. The majority (76.2%) of the studies retrieved had been published between 2010 and 2018. Sampling was done at the

point of retail/trader (57.6%), processing plant (3.8%), or source/farm/point of harvest/manufacture (29.6%). Nine percent of the studies did not specify the point at which sampling was conducted.

The distribution of the studies was varied. Notably, the following states Kebbi, Kwara, Adamawa, Yobe, Zamfara, Jigawa, Kogi, and Katsina were not represented in the reviewed literature. The selected studies covered 28 of the 36 federal states of Nigeria.

Beverage types reported in the review

The most common beverages studied ($n=787$) were sachet water (14.9%), borehole/well water (13.9%), cereal-based beverages (12.1%), raw/fresh milk (8.3%), and *nono/nunu* (7.2%). Other beverage types included river/canal/stream water, processed nonalcoholic beverages, fruit juice, traditional alcoholic beverages, soya milk/tigernut, bottled water, tap/piped water, traditional nonalcoholic drinks, processed milk, processed alcoholic beverages, and rainwater (Table 1).

Hazards reported in the studied beverages

The 5 main hazard groups reported in this review include bacteria, fungi, parasites, biotoxins, and chemical contaminants (a total of 754 records). The most common hazards, or hazard proxies, studied were bacterial (483 records, 64.1%) and chemical contaminants (179 records, 23.7%). A few studies on fungi (55 records, 7.3%), toxins (24 records, 3.2%), and parasites (13 records, 1.7%) were identified. Thirty-three records did not specify the identity of the hazard. The most often studied bacterial genus ($n=483$) was *Escherichia* spp. (16.1%, six studies had negative findings), *Staphylococcus* spp. (14.3%, one study had a negative finding), *Salmonella* spp. (11%, two studies had negative findings), coliforms (8.7%, two studies had negative findings), and *Klebsiella* spp. (6.6%, one study had negative findings). The common fungal and parasitic contaminants studied and reported were as follows ($n=55$; $n=13$, respectively): *Aspergillus* spp. (27.3%), *Penicillium* spp. (14.5%), *Rhizopus* spp. (12.7%), *Giardia* spp. (38.5%), *Entamoeba* spp. (30.8%), and *Ascaris* spp. (23.1%).

TABLE 1. NUMBER OF STUDIES INVESTIGATING THE CONTAMINATION OF SPECIFIC BEVERAGES (TOTAL 787)

| Name of beverage | Number of studies identified (%) | Name of beverage | Number of studies identified (%) |
|-------------------------------------|----------------------------------|--|----------------------------------|
| Sachet water | 117 (14.9) | Yoghurt | 33 (4.2) |
| Borehole/well water | 109 (13.9) | Fermented milk ^a | 25 (3.2) |
| Cereal-based beverage ^b | 95 (12.1) | Soya milk/tigernut | 18 (2.3) |
| Raw/fresh milk | 65 (8.3) | Bottled water | 18 (2.3) |
| <i>Nono/nunu</i> | 57 (7.2) | Tap/piped water | 16 (2) |
| Zobo | 46 (5.8) | Traditional nonalcoholic drinks ^c | 14 (1.8) |
| River/Canal/Stream | 43 (5.5) | Processed milk | 9 (1.1) |
| Processed nonalcoholic ^d | 35 (4.4) | Processed alcoholic ^e | 9 (1.1) |
| Fruit juice | 35 (4.4) | Rainwater | 8 (1) |
| Traditional alcoholic ^f | 35 (4.4) | | |

^aFor example, Kesham, Kindrimo, Manshanu, Wara.

^bKunu, Kunun-zaki, Kunu-aya, Akamu, Ogi.

^cFor example, herbal teas.

^dCanned and noncanned beverages, soft drinks.

^eFor example, beer.

^fFor example, Burukutu, Gin Ufop, Pito.

TABLE 2. HAZARDS REPORTED IN DIFFERENT TYPES OF BEVERAGES IN NIGERIA BETWEEN THE YEARS 2000–2020

| Name/type of beverage | Hazard group | Name of pathogens (genus)/chemicals | Reference |
|--|--------------|---|---|
| Cereal-based beverages (Kunu, Kunun-zaki, Kunu-aya, Akamu, Ogi) Kunu/Kunun-zaki/Kunu-aya is a Cereal-based nonalcoholic fermented beverage, produced by steeping of sorghum, millet, or maize, wet milling, sieving, and partial gelatinization of the slurry (Ndulaka <i>et al.</i> , 2014). Akamu is a porridge made from fermented maize, millet, or sorghum. It is used for weaning infants and taken as breakfast (Nwokoro and Chukwu, 2012) and Ogi is a fermented cereal pudding (Olasupo <i>et al.</i> , 2002) | Bacterial | <i>Bacillus</i> spp., <i>Citrobacter</i> spp., <i>Coliforms</i> , <i>Enterobacter</i> spp., <i>Enterococcus</i> spp., <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Lactobacillus</i> spp., <i>Listeria</i> spp., <i>Proteus</i> spp., <i>Pseudomonas</i> spp., <i>Shigella</i> spp., <i>Salmonella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Olasupo <i>et al.</i> (2002), Umoh <i>et al.</i> (2004), Nwachukwu <i>et al.</i> (2009), Ikpoh <i>et al.</i> (2013), Makut <i>et al.</i> (2013), Aboh and Oladosu (2014), Umaru <i>et al.</i> (2014), Zumbes <i>et al.</i> (2014), Etang <i>et al.</i> (2017), Katuka <i>et al.</i> (2018), Ogodo <i>et al.</i> (2018), Popoola <i>et al.</i> (2019) |
| | Fungal | <i>Aspergillus</i> spp., <i>Candida</i> spp., <i>Fusarium</i> spp., <i>Penicillium</i> spp., <i>Saccharomyces</i> spp., <i>Rhizopus</i> spp. | Ejiogun <i>et al.</i> (2010), Ikpoh <i>et al.</i> (2013), Aboh and Oladosu (2014), Etang <i>et al.</i> (2017) |
| | Chemical | Chromium, iron, lead, zinc | Bakare-Odunola and Mustapha (2014), Maigari <i>et al.</i> (2016) |
| Fermented milk (Kesham, Kindrimo, Manshanu, Wara) | Bacterial | Coliforms, <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Olasupo <i>et al.</i> (2002), Ogbonna <i>et al.</i> (2012), Karshima <i>et al.</i> (2013), Makut <i>et al.</i> (2014b), Umaru <i>et al.</i> (2014), Fowoyo (2016), Musa <i>et al.</i> (2017), Aliyu <i>et al.</i> (2018, 2020) |
| Nono/Nunu Nono/Nunu is crude cultured whole milk (Onyinye <i>et al.</i> , 2020). | Bacterial | <i>Bacillus</i> spp., coliforms, <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Mycobacterium</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Olasupo <i>et al.</i> (2002), Ofukwu <i>et al.</i> (2008), Okonkwo (2011), Yabaya <i>et al.</i> (2012), Karshima <i>et al.</i> (2013), Reuben and Owuna (2013), Agada <i>et al.</i> (2014), Egwaikhede <i>et al.</i> (2014), Ivbade <i>et al.</i> (2014), Makut <i>et al.</i> (2014a), Enem <i>et al.</i> (2015), Enabulele and Nwankiti (2016), Fowoyo (2016), Okpo <i>et al.</i> (2016), Usman and Mustapha (2016), Musa <i>et al.</i> (2017), Dafur <i>et al.</i> (2018), Onioshun (2018), Yakubu <i>et al.</i> (2018), Aliyu <i>et al.</i> (2020) |
| | Fungal | <i>Trichoderma</i> spp., <i>Aspergillus</i> spp., <i>Mucor</i> spp., <i>Candida</i> spp. | Okonkwo (2011), Egwaikhede <i>et al.</i> (2014), Dafur <i>et al.</i> (2018) |
| | Chemical | Antimicrobial residues | Okonkwo (2011), Onyinye <i>et al.</i> (2020) |
| Processed milk (pasteurized milk) | Bacterial | <i>Klebsiella</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Umaru <i>et al.</i> (2014), Dayok <i>et al.</i> (2019) |
| Raw/fresh milk | Chemical | Antimony, mercury, tin | Roberts and Orisakwe (2011) |
| | Bacterial | Coliforms (not further described), <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Lactobacillus</i> spp., <i>Listeria</i> spp., <i>Micrococcus</i> spp., <i>Mycobacterium</i> spp., <i>Proteus</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Campylobacter</i> spp., <i>Cellulomonas</i> spp., <i>Citrobacter</i> spp., <i>Streptococcus</i> spp., <i>Xanthomonas</i> spp., <i>Bacillus</i> spp. | Ofukwu <i>et al.</i> (2008), Cadmus <i>et al.</i> (2010), Salihu <i>et al.</i> (2010), Adesina <i>et al.</i> (2011), Ogbonna <i>et al.</i> (2012), Enurah <i>et al.</i> (2013), Karshima <i>et al.</i> (2013), Agada <i>et al.</i> (2014), Ivbade <i>et al.</i> (2014), Makut <i>et al.</i> (2014a), Oluduro <i>et al.</i> (2014), Umaru <i>et al.</i> (2014), Enem <i>et al.</i> (2015), Mailafia <i>et al.</i> (2017), Onioshun (2018), Yakubu <i>et al.</i> (2018), Dayok <i>et al.</i> (2019), Aliyu <i>et al.</i> (2020) |
| | Chemical | Aflatoxin M1&M2, antimicrobial residues | Chilaka <i>et al.</i> (2018), Onyinye <i>et al.</i> (2020) |

(continued)

TABLE 2. (CONTINUED)

| Name/type of beverage | Hazard group | Name of pathogens (genus)/chemicals | Reference |
|---|--------------|---|---|
| Yoghurt | Bacterial | <i>Aeromonas</i> spp., <i>Bacillus</i> spp., <i>Clostridium</i> spp., Coliforms, <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Lactobacillus</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Nwagu and Amadi (2010), Mbaeyi-Nwaoha and Egbuche (2012), Makut <i>et al.</i> (2014b), Oluduro <i>et al.</i> (2014), Umaru <i>et al.</i> (2014), Chukwu <i>et al.</i> (2015), Enem <i>et al.</i> (2015), Sunday <i>et al.</i> (2016), Usman and Mustapha (2016) |
| | Fungal | <i>Fusarium</i> spp., <i>Geotrichum</i> spp., <i>Neurospora</i> spp., <i>Penicillium</i> spp., <i>Aspergillus</i> spp., <i>Absidia</i> spp. | Sunday <i>et al.</i> (2016) |
| Soya milk/tigernut | Bacterial | <i>Bacillus</i> spp., Coliforms, <i>Escherichia</i> spp., <i>Lactobacillus</i> spp., <i>Salmonella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Brooks <i>et al.</i> (2003), Anagu <i>et al.</i> (2015), Ntukidem <i>et al.</i> (2020) |
| | Fungal | <i>Aspergillus</i> spp., <i>Rhizopus</i> spp. | Brooks <i>et al.</i> (2003), Ntukidem <i>et al.</i> (2020) |
| Processed alcoholic beverages (Beer) | Chemical | Aluminum, cadmium, copper, iron, lead, zinc | Udota and Umoudofia (2011), Jegede <i>et al.</i> (2016), Okareh <i>et al.</i> (2018) |
| Processed nonalcoholic (canned and noncanned beverages, soft drinks) | Chemical | Antimony, arsenic, cadmium, chromium, copper, fluoride, iron, lead, manganese, mercury, nickel, tin, zinc | Maduabuchi <i>et al.</i> (2007, 2008), Roberts and Orisakwe (2011), Adepoju-Bello <i>et al.</i> (2012), Godwill <i>et al.</i> (2015), Magomya <i>et al.</i> (2015), Jegede <i>et al.</i> (2016), Ani <i>et al.</i> (2020) |
| | Fungal | <i>Mucor</i> spp., <i>Mycelia</i> spp., <i>Aspergillus</i> spp., <i>Penicillium</i> spp., <i>Rhizopus</i> spp., <i>Saccharomyces</i> spp., <i>Schizosaccharomyces</i> spp. | Oyetunji (2006) |
| Traditional alcoholic drinks (Burukutu, Gin Ufofop, Pito) | Chemical | Antimony, arsenic, cadmium, nickel, chromium, copper, fluoride, iron, lead, manganese, mercury, tin, zinc | Udota and Umoudofia (2011), Chilaka <i>et al.</i> (2018) |
| | Fungal | <i>Saccharomyces</i> spp., <i>Aspergillus</i> spp., <i>Mucor</i> spp., <i>Schizosaccharomyces</i> spp., <i>Mycelia</i> spp., <i>Rhizopus</i> spp., <i>Penicillium</i> spp. | Olaniyi and Akinyele (2020) |
| | Toxins | Acetyl-deoxynivalenol, fumonisins B1, B2, B3, B4 and zearalenone | Chilaka <i>et al.</i> (2018) |
| Traditional nonalcoholic drinks (herbal tea) | Bacterial | <i>Escherichia</i> spp., <i>Bacillus</i> spp., <i>Klebsiella</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> spp., <i>Staphylococcus</i> spp. | Omogbai and Ikenebomeh (2013) |
| | Fungal | <i>Aspergillus</i> spp., <i>Fusarium</i> spp., <i>Penicillium</i> spp., <i>Rhizopus</i> spp., <i>Serratia</i> spp. | Omogbai and Ikenebomeh (2013) |
| Zobo Zobo is a hot water extract of Hibiscus sabdariffa (Ndulaka <i>et al.</i> , 2014) | Parasitic | <i>Trichuris</i> spp., <i>Giardia</i> spp., <i>Ascaris</i> spp., <i>Entamoeba</i> spp., <i>Balantidium</i> spp. | Ekwunife <i>et al.</i> (2014) |
| | Fungal | <i>Fusarium</i> spp., <i>Aspergillus</i> spp., <i>Penicillium</i> spp., <i>Rhizopus</i> spp., <i>Saccharomyces</i> spp. | Oku <i>et al.</i> (2018), Onuorah and Odibo (2018) |
| | Chemical | Zinc, lead, chromium, iron | Bakare-Odunola and Mustapha (2014), Anagu <i>et al.</i> (2015), Maigari <i>et al.</i> (2016) |

(continued)

TABLE 2. (CONTINUED)

| Name/type of beverage | Hazard group | Name of pathogens (genus)/chemicals | Reference |
|--------------------------|--------------|---|--|
| Fruit Juice | Bacterial | <i>Bacillus</i> spp., <i>Clostridium</i> spp., Coliforms, <i>Shigella</i> spp., <i>Corynebacterium</i> spp., <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Lactobacillus</i> spp., <i>Proteus</i> spp., <i>Salmonella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Mbaeyi-Nwaoha and Egbuche (2012), Risiquat (2013), Ejikeugwu <i>et al.</i> (2014), Umaru <i>et al.</i> (2014), Zumbes <i>et al.</i> (2014), Anagu <i>et al.</i> (2015), Ezeigbo <i>et al.</i> (2015) |
| | Bacterial | <i>Acetobacter</i> spp., <i>Alicyclobacillus</i> spp., <i>Bacillus</i> spp., <i>Enterobacter</i> spp., <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Lactobacillus</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp. | Agwa <i>et al.</i> (2014), Maduka <i>et al.</i> (2014), Ogodo <i>et al.</i> (2016), Osopale <i>et al.</i> (2016) |
| | Fungal | <i>Penicillium</i> spp., <i>Rhizopus</i> spp., <i>Saccharomyces</i> spp., <i>Aspergillus</i> spp. | Agwa <i>et al.</i> (2014), Ogodo <i>et al.</i> (2016) |
| Rainwater | Chemical | Zinc, copper, lead | Okeri <i>et al.</i> (2009) |
| | Bacterial | <i>Enterobacter</i> spp., <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Proteus</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Streptococcus</i> spp., <i>Vibrio</i> spp. | Charity <i>et al.</i> (2012) |
| River/Canal/Stream water | Bacterial | Coliforms, <i>Enterobacter</i> spp., <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Proteus</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp., <i>Vibrio</i> spp. | Iroegbu <i>et al.</i> (2000), Shittu <i>et al.</i> (2008), Charity <i>et al.</i> (2012) |
| Sachet water | Chemical | Lead, nickel, nitrate, nitrite, phosphate, alkalinity, cadmium, chromium, copper, zinc | Duruibe <i>et al.</i> (2007), Adefemi and Awokunmi (2010), Nduka <i>et al.</i> (2010), Yahaya <i>et al.</i> (2012) |
| | Chemical | Lead, magnesium, manganese, molybdenum, PPCPs, nickel, selenium, arsenic, bicarbonate, cadmium, bromine, calcium, chloride, chromium, copper, iron, fluoride, iron, zinc | Oboh <i>et al.</i> (2001), Orisakwe <i>et al.</i> (2006), Okeri <i>et al.</i> (2009), Raji <i>et al.</i> (2010), Abua <i>et al.</i> (2012), Ani <i>et al.</i> (2020), Ebele <i>et al.</i> (2020) |
| | Bacterial | <i>Micrococcus</i> spp., <i>Proteus</i> spp., <i>Vibrio</i> spp., <i>Salmonella</i> spp., <i>Pseudomonas</i> spp., <i>Serratia</i> spp., <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp., <i>Bacillus</i> spp., <i>Klebsiella</i> spp., <i>Chromobacterium</i> spp., coliforms <i>Enterobacter</i> spp., <i>Escherichia</i> spp., <i>Aeromonas</i> spp., <i>Flavobacterium</i> spp., <i>Acinetobacter</i> spp., <i>Alcaligenes</i> spp. | Oboh <i>et al.</i> (2001), Ajayi <i>et al.</i> (2008), Ezeugwunne <i>et al.</i> (2009), Olaoye and Onilude (2009), Mudasiru <i>et al.</i> (2011), Ngwai <i>et al.</i> (2010), Akinyemi <i>et al.</i> (2011), Mgbakor <i>et al.</i> (2011), Mbaeyi-Nwaoha and Egbuche (2012), Oluwafemi and Oluwale (2012), Onilude <i>et al.</i> (2013), Isikwue and Chikezie (2014), Ugochukwu <i>et al.</i> (2015), Okunola <i>et al.</i> (2018) |
| Borehole/well water | Parasitic | <i>Ascaris</i> spp., <i>Trichuris</i> spp., <i>Entamoeba</i> spp., <i>Giardia</i> spp., <i>Ancylostoma/Necator</i> spp. | Ekwunife <i>et al.</i> (2010), Omolade <i>et al.</i> (2017) |
| | Bacterial | <i>Proteus</i> spp., <i>Pseudomonas</i> spp., <i>Salmonella</i> , <i>Shigella</i> spp., <i>Staphylococcus</i> spp., <i>Vibrio</i> spp., <i>Streptococcus</i> spp., Coliforms, <i>Klebsiella</i> spp., <i>Flavobacterium</i> spp., <i>Campylobacter</i> spp., <i>Enterobacter</i> spp., <i>Escherichia</i> spp. | Ibe and Okpleny (2005), Duruibe <i>et al.</i> (2007), Shittu <i>et al.</i> (2008), Mudasiru <i>et al.</i> (2011), Raji <i>et al.</i> (2010), Agbalagba <i>et al.</i> (2011), Akinyemi <i>et al.</i> (2011), Eruola <i>et al.</i> (2011), Charity <i>et al.</i> (2012), Bello <i>et al.</i> (2013), Onwughara <i>et al.</i> (2013), Ugboma <i>et al.</i> (2013), Aboh <i>et al.</i> (2015), Lovet and Dineebimo (2017) |

(continued)

TABLE 2. (CONTINUED)

| Name/type of beverage | Hazard group | Name of pathogens (genus)/chemicals | Reference |
|-----------------------|--------------|--|--|
| Bottled water | Chemical | Mercury, molybdenum, nickel, nitrate and nitrite, PPCPs, phosphate, potassium, selenium, sodium, zinc, calcium, chromium, copper, fluoride, iron, lead, manganese, alkalinity, arsenic, bromine, cadmium | Duruibe <i>et al.</i> (2007), Adefemi and Awokunmi (2010), Mudasiro <i>et al.</i> (2011), Nduka <i>et al.</i> (2010), Agbalagba <i>et al.</i> (2011), Eruola <i>et al.</i> (2011), Adindu <i>et al.</i> (2012), Bello <i>et al.</i> (2013), Egbinola and Amanambu (2014), Ebele <i>et al.</i> (2020) |
| | Bacterial | <i>Escherichia</i> spp., <i>Klebsiella</i> spp., <i>Pseudomonas</i> spp., <i>Staphylococcus</i> spp., <i>Streptococcus</i> spp., coliforms | Ajayi <i>et al.</i> (2008), Igbeneghu and Lamikanra (2014) |
| Tap/piped water | Chemical | Antimony, tin, zinc, lead, iron, chromium copper, fluoride, manganese, mercury, tin, zinc, PPCPs | Okeri <i>et al.</i> (2009), Roberts and Orisakwe (2011), Ani <i>et al.</i> (2020), Ebele <i>et al.</i> (2020) |
| | Bacterial | Coliforms, <i>Escherichia</i> spp., <i>Salmonella</i> spp., <i>Shigella</i> spp. | Iroegbu <i>et al.</i> (2000), Mudasiro <i>et al.</i> (2011), Akinyemi <i>et al.</i> (2011), Isikwue and Chikezie (2014) |

PPCPs, pharmaceuticals and personal care products.

Commonly studied chemical contaminants ($n=179$) were lead (14.5%, seven reports had negative findings), iron (8.9%, seven studies had negative findings), and chromium (8.4%, six studies had negative findings). The fungal metabolites ($n=24$) investigated included acetyl-deoxynivalenol/deoxynivalenol (33.3%), zearalenone (33.3%, three reports had negative findings), and fumonisin (25%). See Tables 2–5 for more details.

Pooled prevalence estimates

Sufficient data were available to allow estimation of pooled prevalence for four hazards in four different beverage types (as described in Table 6). Key hazard/beverage combinations

with sufficient data (>5 high-quality articles) available to undertake this included *Klebsiella* spp. in sachet water, 40.0% (95% CI 12.4–71.7), $I^2=97.3\%$; *Staphylococcus* spp. in *nono/nunu*, 32.6% (95% CI 14.7–53.8), $I^2=97.5\%$; and *Escherichia* spp. in *nono/nunu*, 30.7% (95% CI 21.9–40.2), $I^2=95.7\%$. *Salmonella* spp. in borehole/well water, 19.8% (95% CI 13.1–27.4), $I^2=58.4\%$ —moderate heterogeneity and *Staphylococcus* spp. in raw/fresh milk, 12.3% (95% CI 6.3–20.0), $I^2=34.22\%$. Heterogeneity based on the I^2 statistic was moderate-considerable in 4/5 hazard/beverage pools. The aggregate summary estimates from these studies are therefore to be interpreted with caution. Only one pool was homogeneous, and that was *Staphylococcus* spp. in raw/fresh milk. Forest plots illustrating the data are shown in Figure 2.

TABLE 3. NUMBER OF STUDIES INVESTIGATING BACTERIAL CONTAMINATION OF SPECIFIC BEVERAGES (TOTAL 483)

| Bacteria (genus) | Number of studies (%) | Bacteria (genus) | Number of studies (%) | Bacteria (genus) | Number of studies (%) | Bacteria (genus) | Number of studies (%) |
|----------------------------|-----------------------|------------------------------|-----------------------|-----------------------------|-----------------------|-----------------------------|-----------------------|
| <i>Escherichia</i> spp. | 78 (16.1) | <i>Shigella</i> spp. | 24 (5) | <i>Micrococcus</i> spp. | 5 (1) | <i>Enterococcus</i> spp. | 2 (0.4) |
| <i>Staphylococcus</i> spp. | 69 (14.3) | <i>Proteus</i> spp. | 21 (4.3) | <i>Campylobacter</i> spp. | 4 (0.8) | <i>Aeromonas</i> spp. | 2 (0.4) |
| <i>Salmonella</i> spp. | 53 (11) | <i>Enterobacter</i> spp. | 14 (2.9) | <i>Citrobacter</i> spp. | 4 (0.8) | <i>Alcaligenes</i> spp. | 1 (0.2) |
| Coliforms | 42 (8.7) | <i>Alicyclobacillus</i> spp. | 12 (2.5) | <i>Aeromonas</i> spp. | 4 (0.8) | <i>Acetobacter</i> spp. | 1 (0.2) |
| <i>Klebsiella</i> spp. | 32 (6.6) | <i>Lactobacillus</i> spp. | 9 (1.9) | <i>Chromobacterium</i> spp. | 3 (0.6) | <i>Acinetobacter</i> spp. | 1 (0.2) |
| <i>Streptococcus</i> spp. | 26 (5.4) | <i>Listeria</i> spp. | 7 (1.4) | <i>Clostridium</i> spp. | 3 (0.6) | <i>Xanthomonas</i> spp. | 1 (0.2) |
| <i>Pseudomonas</i> spp. | 25 (5.2) | <i>Vibrio</i> spp. | 7 (1.4) | <i>Serratia</i> spp. | 3 (0.6) | <i>Corynebacterium</i> spp. | 1 (0.2) |
| <i>Bacillus</i> spp. | 25 (5.2) | <i>Mycobacterium</i> spp. | 6 (1.2) | <i>Flavobacterium</i> spp. | 2 (0.4) | | |

TABLE 4. NUMBER OF STUDIES IDENTIFIED INVESTIGATING PARASITIC (TOTAL 13) AND FUNGAL (TOTAL 55) CONTAMINATION OF SPECIFIC BEVERAGES

| Parasite (genus) | Number of studies (%) | Fungus (genus) | Number of studies (%) | Fungus (genus) | Number of studies (%) |
|-------------------------|-----------------------|---------------------------|-----------------------|---------------------------------|-----------------------|
| <i>Giardia</i> spp. | 5 (38.5) | <i>Aspergillus</i> spp. | 15 (27.3) | <i>Mucor</i> spp. | 2 (3.6) |
| <i>Entamoeba</i> spp. | 4 (30.8) | <i>Penicillium</i> spp. | 8 (14.5) | <i>Absidia</i> spp. | 1 (1.8) |
| <i>Ascaris</i> spp. | 3 (23.1) | <i>Rhizopus</i> spp. | 7 (12.7) | <i>Geotrichum</i> spp. | 1 (1.8) |
| <i>Trichuris</i> spp. | 2 (15.4) | <i>Fusarium</i> spp. | 6 (10.9) | <i>Mycelia</i> spp. | 1 (1.8) |
| <i>Balantidium</i> spp. | 1 (7.7) | <i>Saccharomyces</i> spp. | 6 (10.9) | <i>Neurospora</i> spp. | 1 (1.8) |
| <i>Necator</i> spp. | 1 (7.7) | <i>Candida</i> spp. | 5 (9.1) | <i>Schizosaccharomyces</i> spp. | 1 (1.8) |
| | | | | <i>Trichoderma</i> spp. | 1 (1.8) |

Discussion

This review identified beverage types consumed in Nigeria ranging from traditional, nonindustrial, to processed alcoholic and nonalcoholic drinks. However, the five most studied beverages were sachet water, borehole/well water, cereal-based beverages, raw/fresh milk, and *nono/nunu*. Sachet and borehole water are important sources of potable drinking water in Nigeria. The affordability and availability of sachet water make it the primary source of potable water for most of the Nigerian populace (Izah *et al.*, 2016). A survey conducted by Dada and Awotunde (2017) noted a consumer preference for home-prepared beverages as opposed to carbonated drinks. They reported a preference for fruit juice and *kunun-zaki* (cereal-based beverage). *Nono* constitutes part of the crucial staple food for the nomadic Fulani, of Northern Nigeria, and its consumption, together with that of raw milk and milk by-products, is popular among the rural and urban population, a preference that is partly attributed to a belief that it is more nutritious than pasteurized milk products (Egwaikhide *et al.*, 2014).

The spatial assessment revealed an unequal distribution of studies on beverage-associated hazards in Nigeria. This difference may suggest different research priorities by research institutions within these states. It could also imply inadequate resource allocation, to support research, by the states. This is likely the case in neighboring countries given the report by the Global Food Safety Partnership, which highlights the fact that investments that support food safety work are lacking in

sub-Saharan Africa, especially in the dominant informal markets (Global Food Safety Partnership, 2019). The paucity of information on the hazards in beverages from Adamawa, Yobe, Jigawa, Katsina, Kwara, Kogi, Kebbi, and Zamfara states may be indicative of underreporting, or reduced research prioritization in the areas. Notable is that each state in Nigeria has at least two universities (Mogaji, 2019). With the growing interest in food safety, regionally and globally, it is expected that more research will be undertaken to generate evidence on current gaps and effects of interventions.

It was found that much of the literature identified, which was otherwise eligible for review, was deemed of poor quality (130/256) and therefore excluded from this literature review, indicating a potential capacity gap in research design and scientific writing. Limitations in resources, infrastructure, and training have been highlighted as barriers to the generation of high-quality scientific publications in Nigeria and several other African countries (Ajao and Ugwu, 2011; Kumwenda *et al.*, 2017).

This review identified studies on a wide range of biological hazards with records on bacterial hazards constituting the bulk (64%) with genera *Escherichia*, *Staphylococcus*, *Salmonella*, and *Klebsiella* being the most common bacterial pathogens studied. This agrees with a review in selected African countries where *Escherichia coli*, *Salmonella* spp., and *Staphylococcus aureus* were the most frequently studied pathogens in ready-to-eat foods and beverages in the selected studies (Paudyal *et al.*, 2017). These pathogens have been widely associated with foodborne outbreaks and sporadic cases of foodborne toxicity in humans (Ivbade *et al.*, 2014).

TABLE 5. NUMBER OF STUDIES IDENTIFIED INVESTIGATING CHEMICAL (TOTAL 179) AND MYCOTOXIN (TOTAL 24) CONTAMINATION OF BEVERAGES

| Chemical | Number of studies (%) | Chemical | Number of studies (%) | Chemical | Number of studies (%) | Toxin | Number of studies (%) |
|-----------------|-----------------------|------------|-----------------------|------------------------|-----------------------|--|-----------------------|
| Lead | 26 (14.5) | Fluoride | 5 (2.8) | Tin | 3 (1.7) | Acetyl-deoxynivalenol/ deoxynivalenol | 8 (33.3) |
| Iron | 16 (8.9) | Mercury | 5 (2.8) | Alkalinity | 3 (1.7) | Zearalenone | 8 (33.3) |
| Chromium | 15 (8.4) | Antimony | 3 (1.7) | Aluminum | 2 (1.1) | Fumonisin | 6 (25) |
| Copper | 13 (7.3) | Carbonate | 3 (1.7) | Antimicrobial residues | 2 (1.1) | Aflatoxin | 2 (8.3) |
| Zinc | 12 (6.7) | Bromine | 3 (1.7) | Chloride | 2 (1.1) | | |
| Cadmium | 11 (6.1) | Calcium | 3 (1.7) | Magnesium | 2 (1.1) | | |
| Manganese | 10 (5.6) | Molybdenum | 3 (1.7) | Potassium | 1 (0.6) | | |
| Nickel | 10 (5.6) | PPCPs | 3 (1.7) | Sodium | 1 (0.6) | | |
| Arsenic | 9 (5) | Phosphate | 3 (1.7) | | | | |
| Nitrate/Nitrite | 7 (3.9) | Selenium | 3 (1.7) | | | | |

PPCPs, pharmaceuticals and personal care products.

TABLE 6. POOLED PREVALENCE ESTIMATES OF SELECT BIOLOGICAL HAZARDS IN BEVERAGES

| Hazard | Beverage | Study | Sample size | Prevalence (%) | 95% CI | Weight (%) random effects model | | |
|----------------------------|---------------------|------------------------------------|------------------------|----------------------------------|------------------------|------------------------------------|--------|--|
| <i>Klebsiella</i> spp. | Sachet water | Ajayi <i>et al.</i> (2008) | 78 | 80.0 | 69.4–88.2 | 14.58 | | |
| | | Ajayi <i>et al.</i> (2008) | 30 | 89.0 | 72.2–97.4 | 14.21 | | |
| | | Mgbakor <i>et al.</i> (2011) | 24 | 29.2 | 12.6–51.1 | 14.07 | | |
| | | Oluwafemi and Oluwole (2012) | 100 | 2.0 | 0.2–7.0 | 14.64 | | |
| | | Olaoye and Onilude (2009) | 92 | 13.0 | 6.9–21.7 | 14.62 | | |
| | | Ugochukwu <i>et al.</i> (2015) | 20 | 40.0 | 19.1–63.9 | 13.94 | | |
| | | Ugochukwu <i>et al.</i> (2015) | 20 | 40.0 | 19.1–63.9 | 13.94 | | |
| | | Pooled prevalence (random effects) | 364 | 40.0 | 12.4–71.7 | 100.00 | | |
| | | | Test for heterogeneity | | | | | |
| <i>Q</i> | | | 225.7357 | <i>I</i> ² | 97.34% | | | |
| | | | (inconsistency) | | | | | |
| DF | | | 6 | 95% CI for <i>I</i> ² | | 96.05–98.21 | | |
| Significance level | | | <i>p</i> < 0.0001 | | | | | |
| <i>Salmonella</i> spp. | Borehole/well water | Akinyemi <i>et al.</i> (2011) | 37 | 10.8 | 3.0–25.4 | 13.90 | | |
| | | Akinyemi <i>et al.</i> (2011) | 37 | 13.5 | 4.5–28.8 | 13.90 | | |
| | | Akinyemi <i>et al.</i> (2011) | 37 | 24.3 | 11.8–41.2 | 13.90 | | |
| | | Akinyemi <i>et al.</i> (2011) | 37 | 21.6 | 9.8–38.2 | 13.90 | | |
| | | Akinyemi <i>et al.</i> (2011) | 60 | 35.0 | 23.1–48.4 | 16.62 | | |
| | | Akinyemi <i>et al.</i> (2011) | 37 | 8.1 | 1.7–21.9 | 13.90 | | |
| | | Akinyemi <i>et al.</i> (2011) | 37 | 21.6 | 9.8–38.2 | 13.90 | | |
| | | Pooled prevalence (random effects) | 282 | 19.8 | 13.1–27.4 | 100.00 | | |
| | | | Test for heterogeneity | | | | | |
| <i>Q</i> | | | 14.4215 | <i>I</i> ² | 58.40% | | | |
| | | | (inconsistency) | | | | | |
| DF | | | 6 | 95% CI for <i>I</i> ² | | 3.93–81.98 | | |
| Significance level | | | <i>p</i> = 0.0253 | | | | | |
| <i>Staphylococcus</i> spp. | Raw/fresh milk | Dayok <i>et al.</i> (2019) | 17 | 11.8 | 1.5–36.4 | 15.38 | | |
| | | Adesina <i>et al.</i> (2011) | 15 | 26.0 | 7.4–54.4 | 14.08 | | |
| | | Olufemi <i>et al.</i> (2018) | 64 | 14.1 | 6.6–25.0 | 33.12 | | |
| | | Aliyu <i>et al.</i> (2020) | 14 | 7.1 | 0.2–33.9 | 13.40 | | |
| | | Aliyu <i>et al.</i> (2020) | 34 | 2.9 | 0.07–15.3 | 24.02 | | |
| | | Pooled prevalence (random effects) | 144 | 12.3 | 6.3–20.0 | 100.00 | | |
| | | | | | Test for heterogeneity | | | |
| | | <i>Q</i> | | | 6.0808 | <i>I</i> ² | 34.22% | |
| | | | (inconsistency) | | | | | |
| DF | | | 4 | 95% CI for <i>I</i> ² | | 0.0–75.20 | | |
| Significance level | | | <i>p</i> = 0.1932 | | | | | |
| <i>Escherichia</i> spp. | Nono/nunu | Okonkwo (2011) | 200 | 28.0 | 21.9–34.8 | 14.90 | | |
| | | Dafur <i>et al.</i> (2018) | 300 | 16.3 | 12.3–21.0 | 14.97 | | |
| | | Fowoyo and Ogunbanwo (2016) | 54 | 88.9 | 77.4–95.8 | 14.31 | | |
| | | Usman and Mustapha (2016) | 140 | 2.9 | 0.8–7.1 | 14.80 | | |
| | | Usman and Mustapha (2016) | 140 | 13.6 | 8.4–20.4 | 14.80 | | |
| | | Yabaya <i>et al.</i> (2012) | 10 | 100.0 | 69.2–100.0 | 11.77 | | |
| | | Aliyu <i>et al.</i> (2020) | 66 | 6.1 | 1.7–14.8 | 14.45 | | |
| | | Pooled prevalence (random effects) | 910 | 32.6 | 14.7–53.8 | 100.00 | | |
| | | | Test for heterogeneity | | | | | |
| <i>Q</i> | | | 235.8627 | <i>I</i> ² | 97.46% | | | |
| | | | (inconsistency) | | | | | |
| DF | | | 6 | 95% CI for <i>I</i> ² | | 96.24–98.28 | | |
| Significance level | | | <i>p</i> < 0.0001 | | | | | |
| <i>Escherichia</i> spp. | Nono/nunu | Okonkwo (2011) | 200 | 43.0 | 36.0–50.2 | 7.96 | | |
| | | Enabulele and Nwankiti (2016) | 200 | 26.5 | 20.5–33.2 | 7.96 | | |
| | | Enabulele and Nwankiti (2016) | 200 | 24.0 | 18.3–30.5 | 7.96 | | |
| | | Enabulele and Nwankiti (2016) | 200 | 40.5 | 33.6–47.7 | 7.96 | | |
| | | Enabulele and Nwankiti (2016) | 200 | 43.0 | 36.0–50.2 | 7.96 | | |
| | | Enabulele <i>et al.</i> (2015) | 200 | 26.5 | 20.5–33.2 | 7.96 | | |

(continued)

TABLE 6. (CONTINUED)

| Hazard | Beverage | Study | Sample size | Prevalence (%) | 95% CI | Weight (%) random effects model |
|--------|----------|------------------------------------|------------------------|---|-----------|---------------------------------|
| | | Enabulele <i>et al.</i> (2015) | 200 | 24.0 | 18.3–30.5 | 7.96 |
| | | Enabulele <i>et al.</i> (2015) | 200 | 40.5 | 33.6–47.7 | 7.96 |
| | | Enabulele <i>et al.</i> (2015) | 200 | 43.0 | 36.0–50.2 | 7.96 |
| | | Dafur <i>et al.</i> (2018) | 300 | 43.0 | 37.3–48.8 | 8.06 |
| | | Enem <i>et al.</i> (2015) | 127 | 1.6 | 0.2–5.6 | 7.79 |
| | | Yabaya <i>et al.</i> (2012) | 10 | 90.0 | 55.5–99.7 | 4.81 |
| | | Yakubu <i>et al.</i> (2018) | 100 | 2.0 | 0.2–7.0 | 7.67 |
| | | Pooled prevalence (random effects) | 2337 | 30.7 | 21.9–40.2 | 100.00 |
| | | | Test for heterogeneity | | | |
| | | <i>Q</i> | 281.8939 | <i>I</i> ² | | 95.74% |
| | | DF | 12 | (inconsistency) 95% CI for <i>I</i> ² | | 94.10–96.93 |
| | | Significance level | | <i>p</i> < 0.0001 | | |

CI, confidence interval; DF, degrees of freedom.

Characterization of *E. coli* into pathotypes was done in 17/78 records, all reporting enterohemorrhagic *E. coli* in milk and milk products. These pathogens are important causes of diarrhea especially in children younger than 5 years (Havelaar *et al.*, 2015). The pooled prevalence estimate of *Staphylococcus* spp. in raw/fresh milk derived from this study may be useful in complementing other baseline data in milk-safety interventions in Nigeria.

This review revealed a paucity of studies incriminating *Campylobacter* spp., *Vibrio* spp., and *Shigella* spp. as contaminants in beverages yet they are major contributors to foodborne disease burden and have been ranked among the top 10 priority hazards in Africa (Havelaar *et al.*, 2015). *Vibrio cholerae*, the causal agent of cholera, is a significant cause of mortality associated with the consumption of contaminated water. Together with *Shigella* spp. and *Campylobacter* spp., they are common contaminants of water sources in developing countries, stemming from pollution by animal and human feces (Cabral, 2010; Thomas *et al.*, 2020).

Fungi and their metabolites were also reported in beverages. Genera *Aspergillus*, *Penicillium*, and *Rhizopus* were the commonest reported in this review. *Aspergillus* spp. was found to be the most abundant fungus based on pooled prevalence estimates, and high contamination rates were detected in traditional nonalcoholic beverages. Aspergillosis results in symptoms ranging from allergic reactions, saprophytic lung disease, and otomycosis to a systemic disease characterized by immune system failure (Barnes and Marr, 2006). Aspergillosis poses a serious diagnostic and management challenge, hence a threat to the public (Garbino, 2004; Barnes and Marr, 2006). *Rhizopus* spp. and *Penicillium* spp. are important opportunistic pathogens. Their presence additionally introduces the risk of food intoxication and spoilage. Fungal metabolites such as zearalenone, fumonisin, and acetyl-deoxynivalenol were studied and reported. Mycotoxins produce acute to chronic long-term health effects, varying from enteric disease to induction of cancers and immune deficiency (Onuorah and Odibo, 2018). Aflatoxins whose burden was found to be highest in the AFR D subregion (Havelaar *et al.*, 2015) are a particular concern to children who are weaned primarily on cereal (AFB1) and milk-based products (AFM1).

The relatively small number of studies on mycotoxins may present a research gap, considering the vulnerability of the production process of traditional alcoholic beverages to contamination by toxigenic fungi (Olaniyi and Akinyele, 2020). According to Adeloye *et al.* (2019), Nigeria has a high prevalence of alcohol abuse and high consumption of cereal-based alcoholic beverages. This is therefore an area of public health concern. Similarly, the occurrence of parasitic contamination of beverages was scantily documented as a meager 13 records were identified, with all reporting positive findings, despite utilizing relatively insensitive detection techniques (microscopy) (Stensvold and Nielsen, 2012). *Ascaris* spp., *Giardia* spp., and *Entamoeba* spp. were the most common parasites reported from sachet water. Sachet water is a crucial source of drinking water, consumed by over 120 million Nigerians (Izah *et al.*, 2016); this population is potentially at risk of enteric disease caused by parasites, and therefore, quality issues are of significant concern. These parasites have been estimated to account for the ill-health of 68 million people and 765,000 foodborne DALYs in a year (Havelaar *et al.*, 2015). Notably, of the biological hazards identified, no viral pathogens have been reported. This is despite a significant burden from viral hazards, including norovirus (2.5 million DALYs and 35,000 deaths per year, globally), which can be transmitted through contaminated water (Havelaar *et al.*, 2015).

Chemical contaminants, particularly heavy metals, were reported in beverages at levels above the recommended level by the Nigerian regulatory authorities and the WHO. The reported occurrences of heavy metal contamination suggest a growing concern about the potability of beverages consumed in Nigeria. Heavy metals are toxic to animals, humans, and the environment. Due to their toxicity and potential bioaccumulation, serious and mandatory monitoring especially in food should be implemented (Morais *et al.*, 2012). Water is a fundamental element in food processing and a key constituent in the production and preparation process of beverages, including cleaning and sterilization of equipment, and quality issues in water are therefore likely to be transferred to other food. Mogborukor (2012) attributes the decline in potability in Nigeria to environmental deterioration brought about by the increase in population and urbanization. The chemical hazards

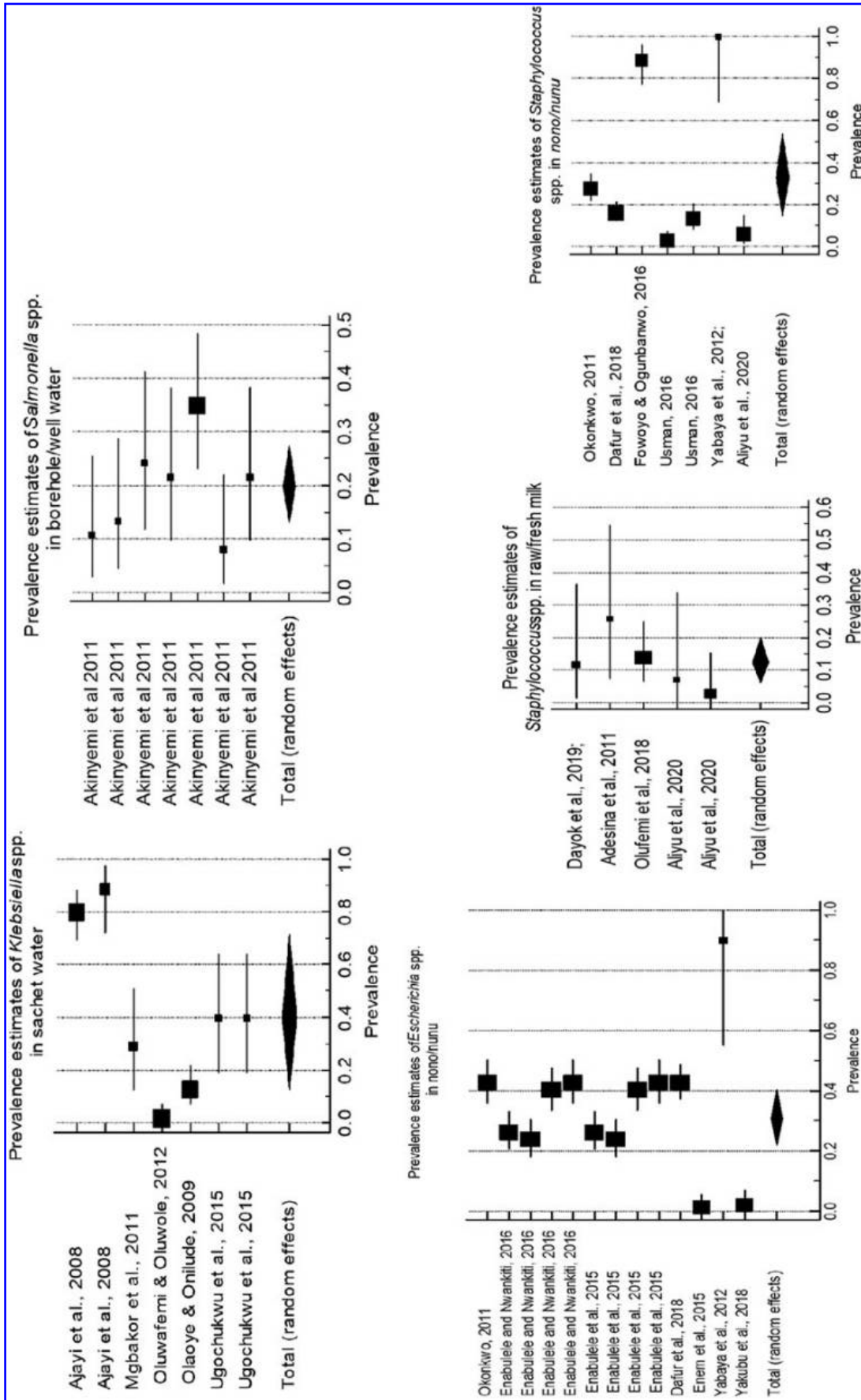


FIG. 2. Forest plots of 5 hazard/beverage combinations showing hazard prevalence estimates in individual studies and the pooled prevalence (random effects model). *Staphylococcus* spp. in raw/fresh milk pool was homogenous. *Salmonella* spp. in borehole/well water, pool had moderate heterogeneity. Heterogeneity was considerable in all the other hazard/beverage combinations as denoted by the non-overlapping confidence intervals of the prevalence estimate in several individual studies in these pools.

outlined here have significant health effects, including brain damage, liver damage, kidney damage, enteric disease, damage to the reproductive system, anemia, cancer, osteopathy, oxidative stress, and oxidation of biological molecules among others (Izah *et al.*, 2016; Engwa *et al.*, 2019).

Food hygiene and safety management systems (FHSM) in many developing African economies are still in their infancy. Its progression has been greatly impeded by the absence or/and underutilization of economic data on the impact of foodborne diseases (Akhtar *et al.*, 2014). Subsequently, there is low prioritization of food safety concerns evidenced by scanty food safety policies, weak supportive legislative environment, and underreporting of foodborne disease outbreaks (Akhtar *et al.*, 2014). Among the components of FHSM is a dynamic and robust risk analysis, which entails hazard identification, characterization, determination of the level of occurrence in food, and exposure assessment among others (Iro *et al.*, 2020). This review outlines deficiencies in quality research outputs at this level. The gaps identified are in alignment with the report by Iro *et al.* (2020) on food safety and management in Nigeria. Iro *et al.* (2020) suggest an integrated approach to Nigeria's food safety concerns, involving the adoption of a Hazard Analysis Critical Control Points (HACCP) system.

The limited capacity of food control laboratories in Africa significantly weakens the surveillance infrastructure both at the national and subnational levels (Akhtar *et al.*, 2014). The main causes include inadequate funding, lack of equipment and personnel, lack of recurrent expenditure to facilitate maintenance of equipment and replenish disposables, and inadequate quality assurance procedures (FAO/WHO, 2005). Developing economies are urged to allocate resources for establishing effective food safety management systems. Accurate, timely, and pertinent information is paramount to compelling policymakers to prioritize investments in food safety systems. Integrated efforts led by research and academic institutes, and encompassing other stakeholders in food safety in a "One Health" framework, should ensure that policymakers receive reliable information on the economic and health implications of food safety management systems and on the measures required to attain quality data (FAO/WHO, 2005; FAO, 2011).

Conclusion

This review has highlighted the presence of several hazards of high importance to public health in commonly consumed beverages in Nigeria and an apparently low investment in the investigation of other significant foodborne hazards known to occur in the African region. The data presented here provide an entry point for future quantitative risk assessments both to determine the level of exposure of the community to these and other hazards, and also for the identification of the most effective mitigation strategies to reduce these risks and improve health outcomes in Nigeria. In addition, the disparity in both the geographic scope and the low consideration of other important hazards may be a useful input in the national foodborne surveillance planning.

Authors' Contributions

D.O.O.: protocol development, methodology, and writing—original draft preparation and editing.

E.K.: protocol development, methodology, and writing—original draft preparation and editing.

L.T.: conceptualization, protocol development, and writing—review and editing.

D.G.: conceptualization, funding acquisition, protocol development, and writing—review and editing.

F.M.: conceptualization, protocol development, and writing—review and editing.

All authors read and approved the final draft of the article.

Acknowledgments

Special thanks to Silvia Alonso for contributing to the development of the study protocol and Elisabetta Lambertini for reviewing the final draft.

Availability of Data and Material

The data are not publicly available but can be provided on request.

Disclosure Statement

No competing financial interests exist.

Funding Information

This study was made possible through support provided by Feed the Future through the United States Agency for International Development (USAID), under the terms of the EatSafe Cooperative Agreement number 7200AA19 CA00010. The opinions expressed herein are those of the author(s) and do not necessarily reflect the views of USAID, the U.S. Agency for International Development, or the U.S. Government. This article is based on a report developed as part of the EatSafe project.

Supplementary Material

Supplementary Data S1

References

- Aboh EA, Giwa FJ, Giwa A. Microbiological assessment of well waters in Samaru, Zaria, Kaduna, State, Nigeria. *Ann Afr Med* 2015;14:32–38.
- Aboh MI, Oladosu P. Microbiological assessment of kunun-zaki marketed in Abuja Municipal Area Council (AMAC) in The Federal Capital Territory (FCT), Nigeria. *Afr J Microbiol Res* 2014;8:1633–1637.
- Abua MA, Iwara AI, Ibor UW, Deekor TD, Ewa EE, Lasisi CJ. A critical assessment of quality status of selected sachet water in Calabar Municipality, Nigeria. *Int J Biosci* 2012;2: 19–26.
- Adefemi S, Awokunmi EE. Determination of physicochemical parameters and heavy metals in water samples from Itaogbolu area of Ondo-State, Nigeria. *Afr J Environ Sci Technol* 2010; 4:145–148.
- Adeloye D, Olawole-Isaac A, Auta A, Dewan MT, Omoyele C, Ezeigwe N, Jacobs W, Mpazanje RG, Harhay MO, Alemu W, Adewole IF. Epidemiology of harmful use of alcohol in Nigeria: A systematic review and meta-analysis. *Am J Drug Alcohol Abuse* 2019;45:438–450.
- Adepoju-Bello AA, Oguntibeju OO, Onuegbu MT, Ayoola GAA, Coker HAB. Analysis of selected metallic impurities in soft drinks marketed in Lagos, Nigeria. *Afr J Biotechnol* 2012;11:4676–4680.

- Adesina K, Oshodi AA, Awoniyi TA. Microbiological assessment of cow milk under traditional management practices in Ado-Ekiti, Nigeria. *Pakistan J Nutr* 2011;10:690–693.
- Adindu RMU, Igboekwe MU, Nnanna L. Groundwater mineralization analysis of Osisioma local government area of Aba, Abia State, Nigeria. *Am J Chem* 2012;2:121–125.
- Agada CA, Adesokan HK, Igwe D, Cadmus SIB. *Mycobacterium africanum* and nontuberculous mycobacteria from fresh milk of pastoral cattle and soft cheese in Oyo State: Implications for public health. *Afr J Med Med Sci* 2014;43:13–20.
- Agbalagba OE, Agbalagba OH, Ononugbo CP, Alao AA. Investigation into the physico-chemical properties and hydrochemical processes of groundwater from commercial boreholes in Yenagoa, Bayelsa State, Nigeria. *Afr J Environ Sci Technol* 2011;5:473–481.
- Agwa O, Ossai-Chidi LN, Ezeani CA. Microbial evaluation of orange fruit juice sold in Port Harcourt, Nigeria. *Am J Food Sci Nutr Res* 2014;1:28–33.
- Ajao O, Ugwu B. Some problems facing scientific medical publications in Nigeria. *J West Afr Coll Surg* 2011;1:31–39.
- Ajayi AA, Sridhar MKC, Adekunle L, Oluwande PA. Quality of packaged waters sold in Ibadan, Nigeria. *Afr J Biomed Res* 2008;11:251–258.
- Akhtar S, Sarker MR, Hossain A. Microbiological food safety: A dilemma of developing societies. *Crit Rev Microbiol* 2014;40:348–359.
- Akinyemi KO, Iwalokun BA, Foli F, Oshodi K, Coker AO. Prevalence of multiple drug resistance and screening of enterotoxin (*stn*) gene in *Salmonella enterica* serovars from water sources in Lagos, Nigeria. *Publ Health* 2011;125:65–71.
- Aliyu Y, Abdullahi IO, Whong CZ, Olalekan BO, Reuben CR. Occurrence and antibiotic susceptibility of methicillin-resistant *Staphylococcus aureus* in fresh milk and milk products in Nasarawa State, North-Central Nigeria. *J Microbiol Antimicrob* 2020;12:32–41.
- Aliyu Y, Reuben RC, Sanj AM, Salawu EM. Occurrence antibiogram of *Staphylococcus aureus* isolated from locally-pasteurised cow milk (kindirmo) sold in parts of Nasarawa Town, Nasarawa State, Nigeria. *Microbiol Res J Int* 2018;23:1–11.
- Anagu L, Emeka O, Moses I, Malachy U, Oli A, Esimone C. Potential spread of pathogens by consumption of locally produced zobo and soya milk drinks in Awka Metropolis, Nigeria. *Br Microbiol Res J* 2015;5:424–431.
- Ani FE, Akaji EA, Uguru NP, Ndiokwelu EM. Fluoride content of commercial drinking water and carbonated soft drinks available in Southeastern Nigeria: Dental and public health implications. *Niger J Clin Pract* 2020;23:65–70.
- Bakare-Odunola MT, Mustapha BK. Identification and quantification of heavy metals in local drinks in Northern Zone of Nigeria. *J Toxicol Environ Health Sci* 2014;6:126–131.
- Barnes PD, Marr KA. Aspergillosis: Spectrum of disease, diagnosis, and treatment. *Infect Dis Clin North Am* 2006;20:545–561.
- Bello OO, Osho A, Bankole SA. Bacteriological and physico-chemical analyses of borehole and well water sources in Ijebu-Ode, Southwestern Nigeria. *IOSR J Pharm Biol Sci* 2013;8:18–25.
- Bown MJ, Sutton AJ. Quality control in systematic reviews and meta-analyses. *Eur J Vasc Endovasc Surg* 2010;40:669–677.
- Brooks AA, Asamudo NU, Udoukpo FC. Microbiological and physico-chemical analysis of soymilk and soyflour sold in Uyo metropolis, Nigeria. *Glob J Pure Appl Sci* 2003;9:457–463.
- Cabral JPS. Water microbiology. Bacterial pathogens and water. *Int J Environ Res Public Health* 2010;7:3657–3703.
- Cadmus SIB, Yakubu MK, Magaji AA, Jenkins AO, van Soelingen D. *Mycobacterium bovis*, but also *M. africanum* present in raw milk of pastoral cattle in north-central Nigeria. *Trop Anim Health Prod* 2010;42:1047–1048.
- Charity EO, Okwouma AC, Emeka IC. Enteric pathogens and diarrhea disease potentials of water sources in Ahiazu Mbaise, Eastern Nigeria. *J Public Health Epidemiol* 2012;4:39–43.
- Chilaka CA, De Boevre M, Atanda OO, De Saeger S. Quantification of *Fusarium mycotoxins* in Nigerian traditional beers and spices using a multi-mycotoxin LC-MS/MS method. *Food Control* 2018;87:203–210.
- Chukwu EE, Ogunsola FT, Nwaokorie FO, Coker AO. Characterization of *Clostridium* species from food commodities and faecal specimens in Lagos State, Nigeria. *West Afr J Med* 2015;34:167–173.
- Dada IO, Awotunde AO. Consumption profile, consumer preference and serving occasions of indigenous non-alcoholic beverages from Nigerian foodstuffs. *Int J Health Sci Res* 2017;7:168–175.
- Dafur GS, Iheukwumere CC, Azua ET, Dafur BS. Evaluation of the microbial quality of 'nono' sold in Mangu Local Government Area of Plateau State, Nigeria. *South Asian J Res Microbiol* 2018;2:1–14.
- Dayok O, Kum FO, Bot TY. Microbial examination of pathogenic bacteria associated with raw and pasteurized milk samples in Shendam L.G.A Plateau State, Nigeria. *World J Innov Res* 2019;7:17–22.
- Deeks JJ, Higgins JPT, Altman DG. Analysing data and undertaking meta-analyses, In: *Cochrane Handbook for Systematic Reviews of Interventions*. Higgins JPT, Thomas J, Chandler J, Cumpston M, Li T, Page MJ, Welch VA, eds. Oxford: Wiley Blackwell, 2019, pp. 241–265.
- Duruibe JO, Ogwuegbu MOC, Ekwurugwu JN. Pollution profiles of non-metallic inorganic and organic pollutants of drinking and potable waters due to mining activities in Ishiagu (Ebonyi State) of Nigeria. *Int J Phys Sci* 2007;2:202–206.
- Ebele AJ, Oluseyi T, Drage DS, Harrad S, Abdallah MAE. Occurrence, seasonal variation and human exposure to pharmaceuticals and personal care products in surface water, groundwater and drinking water in Lagos State, Nigeria. *Emerg Contam* 2020;6:124–132.
- Egbinola CN, Amanambu AC. Groundwater contamination in Ibadan, South-West Nigeria. *SpringerPlus* 2014;3:448.
- Egwaikhede PA, Malu PS, Lawal U, Adelagun RO, Andrew C. Physico-chemical and microbiological analysis of fermented cow milk (nono) consumed within Kaduna Town, North-Western Nigeria. *Food Sci Qual Manage* 2014;29:44–48.
- Ejikeugwu C, Iroha I, Afiukwa N, Nwakaeze E, Oji A, Ilang D. Antibiogram of food-borne pathogens isolated from ready-to-eat foods and zobo drinks sold within and around PRESCO campus of Ebonyi State University (EBSU), Abakaliki, Ebonyi State, Nigeria. *J Toxicol Environ Health Sci* 2014;6:1–4.
- Ejiogu CC, Nnaji AO, Emereibeole EI. Microbiological and physicochemical profiles of kunun-zaki and zoborodo beverages sold in Imo state, Nigeria. *Int J Nat Appl Sci* 2010;6:49–54.
- Ekwunife CA, Okafor CI, Eneanya CI, Ezeunala MN. Human parasitic ova and cyst in local food drinks sold in open markets in Enugu Municipality, South-East, Nigeria. *Bio-scientist* 2014;2:65–69.

- Ekwunife CA, Okafor SO, Ukaga CN, Ozumba NA, Eneanya CI. Parasites associated with sachet drinking water (pure water) in Awka, South-Eastern, Nigeria. *Sierra Leone J Bio Res* 2010;2:23–27.
- Enabulele SA, Nwankiti OO. Shiga toxin (*Stx*) gene detection and verotoxigenic potentials of non-O157 *Escherichia coli* isolated from fermented fresh cow milk (*Nono*) sold in selected cities in Nigeria. *Niger J Basic App Sci* 2016;24:98–105.
- Enem SI, Oboegbulem SI, Nafarnda WD. Detection of verocytotoxigenic *Escherichia coli* O157 serotype in dairy products in Abuja, Nigeria. *Open J Vet Med* 2015;5:224–228.
- Engwa AG, Udoka FP, Nweke NF, Unachukwu NM. Mechanism and health effects of heavy metal toxicity in humans. Available at <https://www.intechopen.com/chapters/64762>, accessed September 9, 2021.
- Enurah LC, Aboaba OO, Nwachukwu SCU, Nwosuh CI. Antibiotic resistant profiles of food (fresh raw milk) and environmental (abattoir effluents) isolates of *Listeria monocytogenes* from the six zones of Nigeria. *Afr J Microbiol Res* 2013;7:4373–4378.
- Eruola AO, Ufoegbune GC, Eruola AO, Awomeso JA, Abuhlimen SA. Analytical assessment of cadmium, lead and iron in hand dug wells of Ilaro, South-Western Nigeria. *Res J Chem Sci* 2011;1:1–5.
- Etang U, Ikon G, Udofia S, Umo A, Udo E, Uyanga F, Ohagim P. Microbiological analyses of kunu drinks locally produced and sold in Calabar, Southern Nigeria. *J Adv Microbiol* 2017;5:1–8.
- Ezeigbo OR, Uhiara S, Nwodu JA, Ekaiko MU. Bacteriological assessment of hawked sorrel drink (zobo drink) in Aba, South-East Nigeria. *Br Microbiol Res J* 2015;5:146–151.
- Ezeugwunne IP, Agbakoba NR, Nnamah NK. The prevalence of bacteria in packaged sachets water sold in Nnewi, South East, Nigeria. *World J Dairy Food Sci* 2009;4:19–21.
- [FAO] Food and Agricultural Organization of the United Nations. Preventing *E. coli* in food. 2011. Available at: www.fao.org/food/food-safety-quality/a-z-index/e-coli0/en, accessed July 9, 2021.
- FAO/WHO. National Food Safety Systems in Africa—A Situational Analysis. 2005. Available at https://www.afro.who.int/sites/default/files/2017-06/fao_who_conf_national_food_safety_africa.pdf, accessed July 9, 2021.
- Fowoyo PT, Ogunbanwo ST. Occurrence and characterisation of coagulase-negative Staphylococci from Nigerian traditional fermented foods. *Food Sci Qual Manage* 2016;50:49–55.
- Garbino DJ. Aspergillosis. 2004. Available at: <https://www.orpha.net/data/patho/GB/uk-Aspergillosis.pdf>, accessed July 9, 2021.
- Gibb HJ, Barchowsky A, Bellinger D, Bolger PM, Carrington C, Havelaar AH, Oberoi S, Zang Y, O’Leary K, Devleeschauwer B. Estimates of the 2015 global and regional disease burden from four foodborne metals—Arsenic, cadmium, lead and methylmercury. *Environ Res* 2019;174:188–194.
- Global Food Safety Partnership. *Food Safety in Africa: Past Endeavors and Future Directions*. Washington, DC: World Bank, 2019.
- Godwill EA, Jane IC, Scholastica IU, Marcellus U, Eugene AL, Gloria OA. Determination of some soft drink constituents and contamination by some heavy metals in Nigeria. *Toxicol Rep* 2015;2:384–390.
- Haddaway NR, Collins AM, Coughlin D, Kirk S. The role of google scholar in evidence reviews and its applicability to grey literature searching. *PLoS One* 2015;10:e0138237.
- Havelaar AH, Kirk MD, Torgerson PR, Gibb HJ, Hald T, Lake RJ, Praet N, Bellinger DC, de Silva NR, Gargouri N, Speybroeck N, Cawthorne A, Mathers C, Stein C, Angulo FJ, Devleeschauwer B, Adegok GO, Afshari R, Alasfoor D, Zeilmaker M. World Health Organization global estimates and regional comparisons of the burden of foodborne disease in 2010. *PLoS Med* 2015;12:1–23.
- Higgins JPT, Savović J, Page MJ, Elbers RG, Sterne JAC. Assessing risk of bias in included studies. *Cochrane Handb Syst Rev Interventions*. 2021. Available at: <https://training.cochrane.org/handbook/current/chapter-08>, accessed July 10, 2021.
- Ibe SN, Okplenyne JI. Bacteriological analysis of borehole water in Uli, Nigeria. *Afr J Zool Environ Biol* 2005;7:116–119.
- Igbeneghu OA, Lamikanra A. The bacteriological quality of different brands of bottled water available to consumers in Ile-Ife, south-western Nigeria. *BMC Res Notes* 2014;7:859.
- Ikpoh IS, Lennox JA, Ekpo IA, Agbo BE, Henshaw EE, Udoekong N. Microbial quality assessment of kunu beverage locally prepared and hawked in Calabar, cross river state, Nigeria. *Glob J Biodivers Sci Manage* 2013;3:58–61.
- Iro OK, Enebeli UU, Iloh GP, Azuama Y, Amadi AN, Amadi CO. Food hygiene and safety management in Nigeria. *Int J Res Sci Innov* 2020;7:101–109.
- Iroegbu CU, Henrietta E, Uwaegbute A, Amazigo UV. Bacteriological quality of weaning food and water in nigerian market bacteriological quality of weaning food and drinking water given to children of market women in Nigeria: Implications for control of diarrhoea. *J Health Popul Nutr* 2000;18:157–162.
- Isikwue MO, Chikezie A. Quality assessment of various sachet water brands marketed in Bauchi metropolis of Nigeria. *Int J Adv Eng Technol* 2014;6:2489–2495.
- Ivbade A, Ojo OE, Dipeolu MA. Shiga toxin-producing *Escherichia coli* O157:H7 in milk and milk products in Ogun State, Nigeria. *Vet Ital* 2014;50:185–191.
- Izah SC, Chakrabarty N, Srivastav AL. A review on heavy metal concentration in potable water sources in Nigeria: Human health effects and mitigating measures. *Expos Health* 2016;8:285–304.
- Jackson D, Turner R. Power analysis for random-effects meta-analysis. *Res Synth Methods* 2017;8:290–302.
- Jaffee S, Henson S, Unnevehr L, Grace D, Cassou E. *The Safe Food Imperative: Accelerating Progress in Low- and Middle-Income Countries*. Agriculture and Food Series 2019. Washington, DC: World Bank, 2020.
- Jegade DO, Oladoye PO, Bamigboye O. Heavy metals assessment in some selected soft and alcoholic drinks in Iwo, Nigeria. *Appl Chem* 2016;95:40838–40841.
- Karshima NS, Pam VA, Bata SI, Dung PA, Paman ND. Isolation of Salmonella species from milk and locally processed milk products traded for human consumption and associated risk factors in Kanam, Plateau State, Nigeria. *J Animal Prod Adv* 2013;3:69–74.
- Katuka YB, Dadah AJ, Yusuf MJ, Uba A. Quinolones resistant enteric bacteria isolated from hawked kunun-zaki in selected parts of Chikun Local Government Area, Kaduna State, Nigeria. *MOJ Toxicol* 2018;4:26–28.
- Kumwenda S, Niang EHA, Orondo PW, William P, Oyinlola L, Bongo GN, Chiwona B. Challenges facing young African scientists in their research careers: A qualitative exploratory study. *Malawi Med J* 2017;29:1–4.
- Lovet TK, Dineebimo B. Bacteriological quality assessment of well water from Amassoma, Bayelsa state, Nigeria. *ASIO J Microbiol Food Sci Biotechnol Innov* 2017;3:9–16.

- Maduabuchi JMU, Adigba EO, Nzegwu CN, Oragwu CI, Okonkwo IP, Orisakwe OE. Arsenic and chromium in canned and non-canned beverages in Nigeria: A potential public health concern. *Int J Environ Res Public Health* 2007;4:28–33.
- Maduabuchi JU, Christine NN, Ese OA, Oragwu CI, Agbo FN, Agbata CA, Ani GC, Orisakwe OE. Iron, manganese and nickel exposure from beverages in Nigeria: A public health concern? *J Health Sci* 2008;54:335–338.
- Maduka HCC, Onuorah OR, Okpogba AN, Ugwu CE, Ogueche PN, Dike CC, Maduka AA. Assessment of some commercial fruit juices commonly consumed in Federal University Of Technology-Owerri (FUTO) by microbiological indices. *IOSR J Pharm Biol Sci* 2014;9:56–58.
- Magomya AM, Yebpella GG, Okpaegbe UC. An Assessment of metal contaminant levels in selected soft drinks sold in Nigeria. *Int J Innov Sci Eng Technol* 2015;2:517–522.
- Maigari AU, Sulaiman MB, Maigari IA. Heavy metals contamination in two popular local drinks consumed in northern Nigeria. *Int J Innov Sci Eng Technol* 2016;3:441–446.
- Mailafia S, Olatunde HO, Okoh G, Chinyere J, Adamu SG, Onyilokwu SA. Microbat™ 24E system identification and antimicrobial sensitivity pattern of bacterial flora from raw milk of apparently healthy lactating cows in Gwagwalada, Nigeria. *J Coast Life Med* 2017;5:356–359.
- Makut MD, Nyam MA, Amapu T, Ahmed A. Antibigram of bacteria isolated from locally processed cow milk products sold in Keffi Metropolis, Nasarawa State, Nigeria. *J Biol Agric Healthc* 2014a;4:19–25.
- Makut MD, Nyam MA, Obiekezie SO, Abubakar, EA. Antibigram of bacteria isolated from Kunun-zaki drink sold in Keffi metropolis. *Am J Infect Dis* 2013;9:71–76.
- Makut MD, Ogbonna A, Dalami H. An assessment of the bacteriological quality of different brands of yoghurt sold in Keffi, Nasarawa State, Nigeria. *J Nat Sci Res* 2014b;4: 19–22.
- Mbaeyi-Nwaoha IE, Egbuche N. Microbiological evaluation of sachet water and street-vended yoghurt and “Zobo” drinks sold in Nsukka metropolis. *Int J Biol Chem Sci* 2012;6:1703–1717.
- Mensah P, Mwamakamba LMC, Nsue-Milang D. Public health and food safety in the WHO African region. *Afr J Food Agric Nutr Dev* 2012;4:14–17.
- Mgbakor C, Ojiegbe GC, Okonko IO, Odu NN, Alli JA, Nwanze JC, Onoh CC. Bacteriological evaluation of some sachet water on sales in Owerri Metropolis, Imo State, Nigeria. *Malay J Microbiol* 2011;7:217–225.
- Mogaji E. Types and location of Nigerian Universities. *Research Agenda Working Papers* 2019;2019:92–103.
- Mogborokor JO. Domestic sewage disposal and quality of water from hand dug wells in Ughelli, Nigeria. *AFRREV STECH Int J Sci Technol* 2012;1:112–125.
- Moher D, Liberati A, Tetzlaff J, Altman DG; The PRISMA Group. Preferred reporting items for systematic reviews and meta-analysis: The PRISMA statement. *PLoS Med* 2009;6: e1000097.
- Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P, Stewart LA, PRISMA-P Group. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015 statement. *Syst Rev* 2015;4:1–9.
- Morais S, Costa FG, Pereira ML. Heavy metals and human health. *Environ Health Emerg Issues Pract* 2012;2012:228–243.
- Mudasiru IOR, Yakubu KEI, Ehiinmidu J. Bacteriological quality of public water sources in Shuni, Tambuwal and Sokoto towns in North-Western Nigeria. *J Pharm Bioresour* 2011;7:55–64.
- Muller O, Krawinkel M. Malnutrition and health in developing countries. *Rev Synth* 2005;3:279–286.
- Musa JA, Dauda KI, Lawan FA, Diyo D, Maina MM, Jauro S. Prevalence and antibiotic sensitivity pattern of Salmonella isolates from milk products and water reservoirs in Maiduguri, North-Eastern Nigeria. *IOSR J Agric Vet Sci* 2017;10: 87–92.
- Nduka JK, Orisakwe OE, Ezenweke LO. Nitrate and nitrite levels of potable water supply in Warri, Nigeria: A public health concern. *J Environ Health* 2010;72:28–31.
- Ndulaka JC, Obasi NE, Omeire GC. Production and Evaluation of Reconstitutible Kunun-Zaki. *Niger Food J* 2014;32: 66–72.
- Ngwai YB, Sounyo AA, Fiabema SM, Agadah GA, Ibeakuzie TO. Bacteriological safety of plastic-bagged sachet drinking water sold in Amassoma, Nigeria. *Asian Pac J Trop Med* 2010;3:555–559.
- [NPC] National Population Commission [Nigeria] and ICF. *Nigeria Demographic and Health Survey 2018*. Abuja, Nigeria, and Rockville, Maryland, USA: NPC and ICF, 2019.
- Ntukidem VE, Edima-Nyah AP, Ndah LS, Nsikak AA. Assessment of microbiological safety and organoleptic properties of tiger-nut (*Cyperus esculentus*) beverage processed locally and sold in Uyo Metropolis of Akwa Ibom State, Nigeria. *Int J Food Nutr Saf* 2020;11:37–50.
- Nwachukwu NC, Orji FA, Amike JI. Isolation and characterization of *Listeria monocytogenes* from kunu, a locally produced beverage marketed in different markets in Abia State of Nigeria. *Aust J Basic Appl Sci* 2009;3:4432–4436.
- Nwagu TN, Amadi EC. Bacteria population of some commercially prepared yoghurt sold in Enugu State, Eastern Nigeria. *Afr J Microbiol Res* 2010;4:984–988.
- Nwokoro O, Chukwu BC. Studies on akamu, a traditional fermented maize food. *Revi Chile Nutri* 2012;39:180–184.
- Obboh G, Adetuyi FC, Akinyosoye FA. Safety evaluation of some packaged potable water in Ondo State, Nigeria. *Nigerian Soc Exp Biol* 2001;1:306–310.
- Ofukwu R, Oboegbulem SI, Akwuobu CA. Zoonotic *Mycobacterium* species in fresh cow milk and fresh skimmed, unpasteurised market milk (nono) in Makurdi, Nigeria: Implications for public health. *J Anim Plant Sci* 2008;1:21–25.
- Ogbonna IO, David AB, Waba JT, Eze PC. Microbiological quality assessments of biradon, kesham and kindrimo: Milk products sold in Maiduguri, Nigeria. *Int J Dairy Sci* 2012;7: 11–19.
- Ogodo AC, Agwaranze DI, Nwaneri CB, Yakubu MN, Hussaini ZJ. Comparative study on the bacteriological quality of kunun-aya sold in Wukari, Nigeria. *Int J Res Stud Microbiol Biotechnol* 2018;4:23–29.
- Ogodo AC, Ugbogu OC, Ekeleme UG, Nwachukwu NO. Microbial quality of commercially packed fruit juices in South-East Nigeria. *J Basic Appl Res* 2016;2:240–245.
- Okareh OT, Oyelakin TM, Ariyo O. Phytochemical properties and heavy metal contents of commonly consumed alcoholic beverages flavoured with herbal extract in Nigeria. *Beverages* 2018;6:3–9.
- Okeri HA, Mmeremikwu AC, Ifeadi AN. Determination of trace metals presence in drinking water and fruit juice in Benin City, Nigeria. *J Appl Biosci* 2009;3:700–702.

- Okonkwo OI. Microbiological analyses and safety evaluation of nono: A fermented milk product consumed in most parts of Northern Nigeria. *Int J Dairy Sci* 2011;6:181–189.
- Okpo NO, Abdullahi IO, Whong CMZ, Ameh J. Occurrence and antibiogram of *Staphylococcus aureus* in dairy products consumed in parts of Kaduna State, Nigeria. *Bayero J Pure Appl Sci* 2016;9:225–229.
- Oku I, Alagoa K, Daworiye P, Izon-ebi B. Microbial content of zobo drink from five different producers within Yenagoa City Bayelsa State, Nigeria. *Int J Adv Sci Res Eng* 2018;4:74–89.
- Okunola OJ, Oba DO, Oranusi SU, Okagbue HI. Data on microbial assessment and physicochemical characteristics of sachet water samples obtained from three factories in Ota, Ogun state, Nigeria. *Data Brief* 2018;19:2445–2451.
- Olaniyi OO, Akinyele JB. Isolation of toxigenic *Aspergillus flavus* and evaluation of aflatoxins in “Burukutu,” sorghum fermented beverage sold in Akure, Nigeria. *J Food Saf Hyg* 2020;5:3882.
- Olaoye OA, Onilude AA. Assessment of microbiological quality of sachet-packaged drinking water in Western Nigeria and its public health significance. *Public Health* 2009;123:729–734.
- Olasupo NA, Smith SI, Akinsinde KA. Examination of the microbial status of selected indigenous fermented foods in Nigeria. *J Food Saf* 2002;22:85–93.
- Oluduro AO, Abike TO, Oriade KD. Phenotypic and molecular characterization of *Salmonella* serotypes in cow raw milk and milk products in Nigeria. *Afr J Biotechnol* 2014;13:3374–3789.
- Olufemi FO, Akinduti PA, Keinde OB, Odunfa OA. Prevalence and antibiogram of methicillin-susceptible *Staphylococcus aureus* (MSSA) isolated from raw milk of asymptomatic cows In Abeokuta, Nigeria. *Alex J Veter Sci* 2018;57:34–40.
- Oluwafemi F, Oluwole ME. Microbiological examination of sachet water due to a cholera outbreak in Ibadan, Nigeria. *Open J Med Microbiol* 2012;2:115–120.
- Omolade O, Zanaib, G. Parasitological evaluation of sachet drinking water in areas of Lagos State, Nigeria. *Electron J Biol* 2017;13:144–151.
- Omogbai BA, Ikenebomeh M. Microbiological characteristics and phytochemical screening of some herbal teas in Nigeria. *Eur Sci J* 2013;9:149–160.
- Onilude AA, Adesina FC, Oluboyede OA, Adeyemi BI. Microbiological quality of sachet packaged water vended in three local governments of Oyo State, Nigeria. *Afr J Food Sci Technol* 2013;4:195–200.
- Onioshun, E. Occurrence and antibiogram of *Salmonella* and *Shigella* species from raw and fermented cow milk (“Nono”) in Zaria and Environs. Available at <https://bit.ly/3yPoDUF>, accessed July 11, 2021.
- Onuorah S, Odibo F. Characterization and identification of fungi in the sorrel beverage (zobo) hawked in Ifite Awka, Anambra State, Nigeria. *Int J Homeopath Nat Med* 2018;4:24–30.
- Onwughara NI, Ajiwe VE, Chima HON, Chima CH. Bacteriological assessment of selected borehole water samples in Umuahia North Local Government Area, Abia State, Nigeria. *J Environ Treat Techn* 2013;1:117–121.
- Onyinye SO, Ezenduka VE, Anaelom NJ. Screening for tylosin and other antimicrobial residues in fresh and fermented (nono) cow milk in Delta state, South-South, Nigeria. *Vet World* 2020;13:458–464.
- Orewole M, Makinde OW, Adekalu KO, Shittu KA. Chemical examination of piped water supply of Ile-Ife in Southwest Nigeria. *Iran J Environ Health Sci Eng* 2007;4:51–56.
- Orisakwe OE, Ogwilo IO, Afonne OJ, Maduabuchi JU, Obi E, Nduka JC. Heavy metal hazards of sachet water in Nigeria. *Arch Environ Occup Health* 2006;61:208–213.
- Osopale BA, Witthuhn CR, Albertyn J, Oguntoyinbo FA. Culture dependent and independent genomic identification of *Alicyclobacillus* species in contaminated commercial fruit juices. *Food Microbiol* 2016;56:21–28.
- Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan-a web and mobile app for systematic reviews. *Syst Rev* 2016;5:1–10.
- Oyetunji TO. Mycological evaluation of a ground cocoa-based beverage. *Afr J Biotechnol* 2006;5:2073–2076.
- Paudyal N, Anihouvi V, Hounhouigan J, Matsheka MI, Sekwati-Monang B, Amoa-Awua W, Atter A, Ackah NB, Mbugua S, Asagbra A, Abdelgadir W, Nakavuma J, Jakobsen M, Fang W. Prevalence of foodborne pathogens in food from selected African countries—A meta-analysis. *Int J Food Microbiol* 2017;249:35–43.
- Popoola OO, Balogun DAA, Bello A. Microbiological quality of some selected akamu samples sold in some areas of Kano Metropolis (A case study of Hotoro, Tarauni and Mariri). *Res J Food Sci Qual Control* 2019;5:8–11.
- R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing, 2018.
- Raji M, Ibrahim Y, Ehinmidu J. Physico-chemical characteristics and heavy metal levels in drinking water sources in Sokoto Metropolis in North-western Nigeria. *J Appl Sci Environ Manage* 2010;14:81–85.
- Reuben RC, Owuna G. Antimicrobial resistance patterns of *Escherichia coli* O157:H7 from Nigerian fermented milk samples in Nasarawa State, Nigeria. *Int J Pharm Sci Invent* 2013;2:38–44.
- Risiquat RO. Bacteriology quality of zobo drinks consumed in some parts of Osun State, Nigeria. *J Appl Sci Environ Manage* 2013;17:113–117.
- Roberts II, Orisakwe OE. Evaluation of potential dietary toxicity of heavy metals in some common Nigerian beverages: A look at antimony, tin and mercury. *QScience Connect* 2011;2:2–10.
- Salihu MD, Junaidu AU, Magaji AA, Rabi ZM. Study of *Campylobacter* in raw cow milk in Sokoto State, Nigeria. *Br J Dairy Sci* 2010;1:1–5.
- Shittu OB, Olaitan JO, Amusa T. Physico-chemical and bacteriological analyses of water used for drinking and swimming purposes in Abeokuta, Nigeria. Analysis of drinking and swimming water in Abeokuta. *Afr J Biomed Res* 2008;11:285–290.
- Stensvold CR, Nielsen HV. Comparison of microscopy and PCR for detection of intestinal parasites in Danish patients supports an incentive for molecular screening platforms. *J Clin Microbiol* 2012;50: 540–541.
- Sunday AN, Chukwuebuka AK, Juliet M, Chidiebere I, Benjamin OC, Gladys AC, Joseph AE, Ada IC, Uche N, Nnamdi U. Microbial assessment of yoghurts sold in Amawbia, Nigeria. *Univ J Microbiol Res* 2016;4:55–58.
- Thomas KM, de Glanville WA, Barker GC, Benschop J, Buza JJ, Cleaveland S, Davis MA, French NP, Mmbaga BT, Prins G, Swai ES, Zadoks RN, Crump JA. Prevalence of *Campylobacter* and *Salmonella* in African food animals and meat: A systematic review and meta-analysis. *Int J Food Microbiol* 2020;315:108382. <https://doi.org/10.1016/j.ijfoodmicro.2019.108382>

- Udota HIJ, Umoudofia SJ. Heavy metal contamination of some selected Nigerian and imported alcoholic drinks. *J Ind Pollut Control* 2011;27:1–4.
- Ugboma AN, Salihu MD, Magaji A, Abubakar M. Prevalence of *Campylobacter* species in ground water in Sokoto, Sokoto State, Nigeria. *Vet World* 2013;6:285–287.
- Ugochukwu S, Giwa FJ, Giwa A. Bacteriological evaluation of sampled sachet water sold in Samaru-Zaria, Kaduna-State, Nigeria. *Nigerian J Basic Clin Sci* 2015;12:1–12.
- Umaru GA, Tukur IS, Akensire UA, Adamu Z, Bello OA, Shawulu AHB, Audu M, Sunkani JB, Adamu G, Adamu NB. Microflora of kunun-zaki and sobo drinks in relation to public health in Jalingo Metropolis, North-Eastern Nigeria. *Int J Food Res* 2014;1:16–21.
- Umoh VJ, Oranusi SU, Kwaga JKP. The public health significance of pathogens isolated from kunun-zaki sold in retail outlets in Zaria, Nigeria. *Nigerian Food J* 2004;22:10–17.
- Usman RZ, Mustapha BM. Isolation and identification of methicillin resistant *Staphylococcus aureus* (MRSA) from traditionally fermented milk “Nono” and yoghurt in Zaria Metropolis, Nigeria. *Int J Compr Lead Res Sci* 2016;2:1–21.
- Yabaya A, Manga SS, Lucy M, Alhassan H. Bacteriological quality of fermented milk sold locally in Samaru and Sabongari Market, Zaria–Nigeria. *Continental J Microbiol* 2012;6:14–18.
- Yahaya A, Adegbe AA, Emurotu JE. Assessment of heavy metal content in the surface water of Oke-Afa Canal Isolo Lagos, Nigeria. *Arch Appl Sci Res* 2012;4:2322–2326.
- Yakubu Y, Shuaibu AB, Ibrahim AM, Hassan UL, Nwachukwu RJ. Risk of Shiga toxigenic *Escherichia coli* O157:H7 infection from raw and fermented milk in Sokoto Metropolis, Nigeria. *J Pathog* 2018;2018:1–5.
- Zumbes JH, Dabo AD, Dakul DA, Afolabi SA, Apiya HS. Enteropathogenic bacterial contamination of some ready to eat foods sold in Jos Metropolis, Nigeria. *Indian J Appl Res* 2014;4:456–458.

Address correspondence to:
Florence Mutua, BVM, MSc, PhD
International Livestock Research Institute
P.O. Box 30709
Nairobi 00100
Kenya

E-mail: f.mutua@cgiar.org